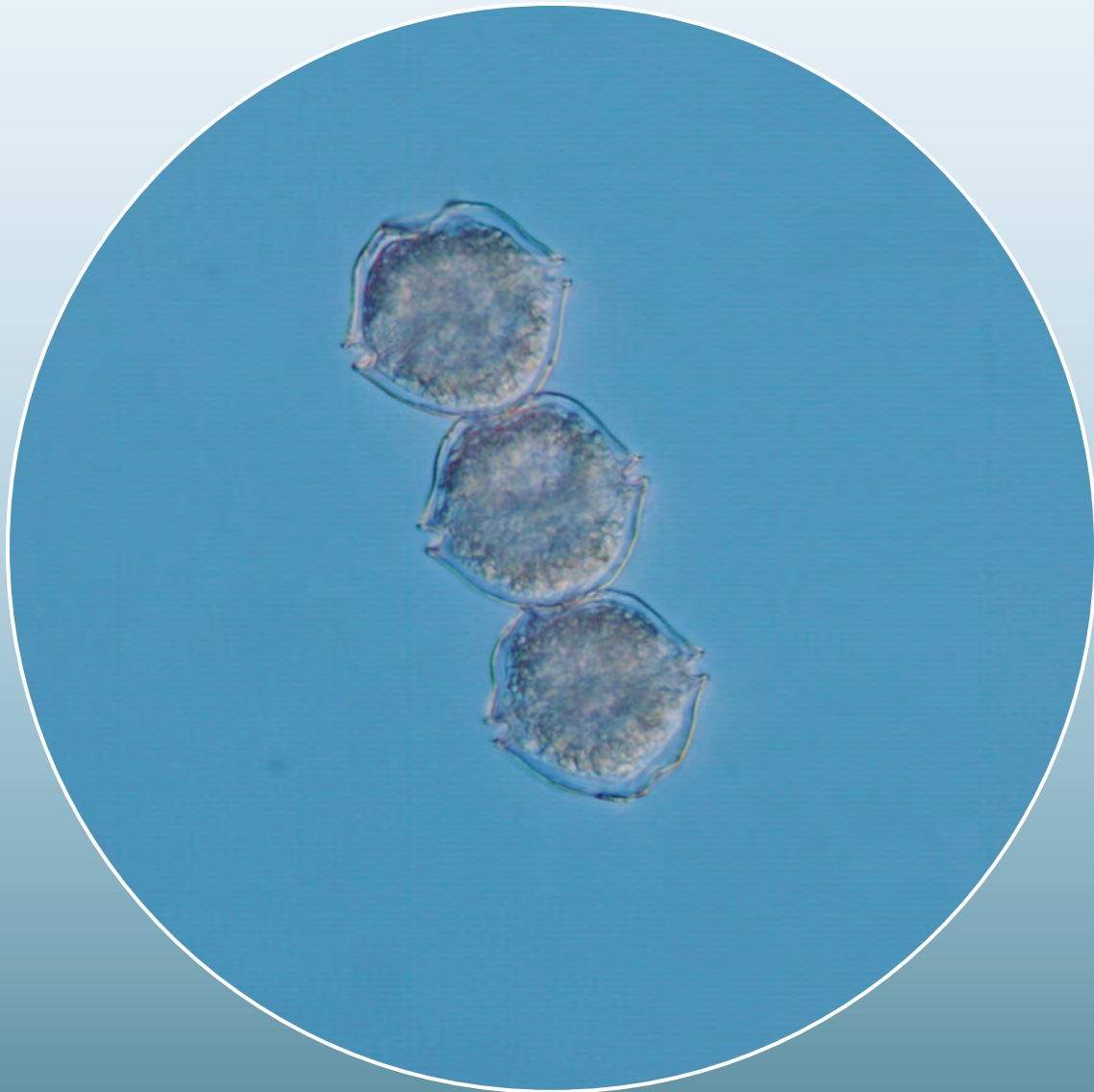


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Exploratory floristics of epiphytic diatoms from
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Exploratory floristics of epiphytic diatoms from Revillagigedo Islands (Mexico)

Florística exploratoria de diatomeas epifitas de Islas Revillagigedo (México)

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ABSTRACT

The feasibility of surveying substrates from remote localities such as Islas Revillagigedo (Mexico) promised both to increase the number of benthic diatom taxa hitherto known from Mexican littorals, and to demystify the endemic potential of the archipelago. Because the surface of rhodophytes harbor a great many diatom taxa, we expected that an exploratory survey using only two specimens of *Laurencia* sp. would provide an approximate estimate of the species richness of epiphytic diatoms living on macroalgae from the archipelago. Our survey yielded 208 diatom taxa (species and varieties) that, although most have been recorded elsewhere, it comprises 52 new additions to the benthic diatom flora for Mexican littorals, including 16 undetermined taxa that are likely new species. The high number of *Mastogloia* taxa, represented by 27 species and varieties, plus several other recorded taxa from the tropics, and the many new records not observed in the Mexican NW, suggests a strong tropical affinity of the diatom taxocenosis that thrive in the Revillagigedo islands.

Key words: Bacillariophyta; diversity; new records, red algae; species richness.

RESUMEN

La posibilidad de revisar sustratos de localidades remotas como las Islas Revillagigedo (México) prometía, tanto incrementar el número de taxones de

diatomeas bentónicas conocidas hasta ahora en litorales mexicanos, como desmitificar el potencial endémico del archipiélago. Dado que la superficie de rodofitas sirve de sustrato a muchas especies de diatomeas, asumimos que una inspección exploratoria en solo dos talos de *Laurencia* sp. proveería un estimado aproximado de la riqueza de especies de diatomeas epifitas de macroalgas del archipiélago. La florística redituó 208 taxones de diatomeas, entre especies y variedades; y, aunque la mayoría han sido registrados en otras localidades, comprende 52 adiciones a la flora de diatomeas bentónicas de litorales mexicanos, incluyendo 16 taxones no determinados que quizá sean nuevas especies. El elevado número de taxones del género *Mastogloia*, representado por 27 especies y variedades, aunado a los varios taxones registrados en localidades tropicales, así como los nuevos registros hasta ahora no observados en el noroeste mexicano, sugieren una fuerte afinidad tropical de la taxocenosis de diatomeas epifitas que habitan en las Islas Revillagigedo.

Palabras clave: Algas rojas; Bacillariophyta; diversidad; nuevos registros; riqueza de especies.

INTRODUCTION.

The high floristic potential of benthic diatoms for Mexican littorals has been recently confirmed. Studies in hitherto unexplored areas and the sur-

veying of new substrata have yielded numerous new records (López-Fuerte *et al.* 2015; Martínez & Siqueiros-Beltrones 2018; Siqueiros Beltrones *et al.* 2017; Siqueiros Beltrones & Martínez 2017) that have enriched the current overall species list for the region (López-Fuerte & Siqueiros Beltrones 2016). This may be related to the great variety of substrata potentially used by benthic diatoms, which includes both live and inert surfaces. Among the former, many animals harbor high diversities of diatoms, but the surfaces of macroalgae provide by far a higher species richness of epiphytic diatoms which surpasses several hundreds of species. In spite of the latter, many macroalgae taxa are still to be surveyed for epiphytic diatom floristics.

In Mexico, most research on the subject has been carried out for the NW region and, although these are scarce there are even fewer for the tropical zones. Consequently, for the southern Gulf of Mexico only one *ex profeso* study on epiphytic diatoms of rhodophytes has been published. This exploratory study yielded 115 epiphytic diatom taxa from six rhodophyte species, that included six new records for Mexican waters (Siqueiros Beltrones & Martínez 2017). Thus, the possibility of observing substrata from remote tropical localities such as Islas Revillagigedo promises an increase in the number of new records of benthic diatoms for the Mexican littorals and the likeliness of new species. Likewise, it opens expectations on the particularities of benthic diatoms taxocoenoses as well demystification of the alleged endemism for the islands as marked by the Comisión Nacional de Areas Naturales Protegidas and the Secretaría de Medio Ambiente y Recursos Naturales, of Mexico (CONANP-SEMARNAT 2015).

Four islands make up the Revillagigedo archipelago, San Benedicto, Socorro, Roca Partida and Clarion (UNESCO 2018). Under the jurisdiction of the state of Colima, Mexico they are located 386 km south of Cabo San Lucas, Baja California Sur. Although on July 2016 the UNESCO declared it World Heritage Site, only recently (CONANP 2017) was there a previous study issued for justifying its declaration as a national park. In spite of the comprehensive character of the above study, much research is yet to be done in the archipelago, inasmuch its remote location contributes to the conservation of multiples habitats and distinct substrata used by benthic fauna and flora that are yet to be described.

A previous report on diatoms from the archipelago (Hanna & Grant 1926) deals with fossil forms. In this study, we focused on epiphytic diatoms living on red macroalgae, inasmuch the surface of macroalgae and particularly rhodophytes have

been observed to harbor many diatom taxa (Siqueiros Beltrones & Argumedo Hernández 2014; Siqueiros Beltrones & Hernández-Almeida 2006; Siqueiros Beltrones & Martínez 2017). The benthic macroalgal flora for Revillagigedo Islands comprises at least 190 taxa, including 123 rhodophytes (Serviere-Zaragoza *et al.* 2007), although as much as 205 taxa (134 rhodophytes) have been identified (León-Tejera *et al.* 1996).

Seasonal variations in the region seem to be determined by the alternating influence of the California Current and the North-equatorial Current, whilst the rest of the year a transition between both states may be observed (Lluch-Cota *et al.* 1994). These environmental characteristics lead us to propose a hypothesis on the biogeographical affinity of the epiphytic diatom taxocoenosis, which in this case promised to be strongly tropical although with important temperate components.

Because the remoteness of the archipelago largely precludes access to the islands habitats in order to carry out a comprehensive sampling that is representative of the target taxa, we hoped that a small sample consisting of two macroalgae specimens collected *ex profeso* would provide an approximate idea of the species richness potential of the rhodophyte epiphytic diatoms for the islands. Thus, our objective was to construct an exploratory floristic list of epiphytic diatoms of rhodophyte macroalgae for the Revillagigedo archipelago that comprised as many taxa as could be surveyed to support further formal taxonomic and hypothesis driven studies on diatom floristics, ecology, and biogeography for the Revillagigedo Archipelago.

MATERIAL AND METHODS.

Two red algae specimens were collected manually by Scuba during the December 27-January 4, 2018 period at the rocky shore of San Benedicto Island (Revillagigedo Archipelago) at 15-25 m depth (Map 1). The specimens were sun-dried, transported in a plastic ziploc bag and identified in the laboratory following Abbot & Hollenberg (1976). Diatoms were brushed off from each specimen of red algae while rinsing with purified water. The resulting sample was placed in a 150-ml test tube and left to settle. Then, the sample precipitate was collected and oxidized with a mixture of commercial alcohol and nitric acid at a ratio of 1(sample): 2 (alcohol): 5 (acid), (Siqueiros Beltrones 2002). The oxidized material was rinsed repeatedly with purified water until it reached a pH \geq 6. For each sample two double permanent slides were mounted using Pleurax (RI=1.7). Identification was done at 1000 \times under an

Olympus CH-2 compound microscope with phase contrast illumination.

Diatoms were identified following Cleve-Euler (1952, 1953a, 1953b, 1955), Desikachary (1988, 1989), Desikachary & Prema (1987), Desikachary *et al.* (1987), Foged (1975, 1978, 1984), Hernández-Almeida *et al.* (2013), Hernández-Almeida and Siqueiros-Beltrones (2008, 2012), Hustedt (1955, 1959, 1961-66), Joon *et al.* (2018), Hein *et al.* (2008), Loir and Novarino (2013), López-Fuerte *et al.* (2010), Moreno *et al.* (1996), Peragallo and Peragallo (1908), Siqueiros-Beltrones (2002, 2006), Siqueiros-Beltrones and Hernández-Almeida (2006), Siqueiros-Beltrones *et al.* (2014), Schmidt *et al.* (1874 -1959), Stepanek *et al.* (2016), Stidolph *et al.* (2012), Van Heurck (1881), Witkowski *et al.* (2000). Taxonomic status was updated according to the AlgaeBase web site (Guiry & Guiry 2018), and Round *et al.* (1990). A catalog of the recorded epiphytic diatoms was assembled with micrographs of specimens taken with a CMOS Konus digital ocular lens microscope at 1000 \times .

RESULTS.

A total of 208 (specific and infra-specific) epiphytic diatom taxa were identified in the samples from both specimens of *Laurencia* collected at Revillagigedo Islands; sixteen of these taxa could not be identified to species level and may represent new taxa. Also, 52 taxa are new additions to the diatom flora of the Mexican coasts and three to the Mexican Pacific coast: *Cocconeis caribensis* Romero et Navarro, *Mastogloia affirmata* (Leudiger-Fortmore) Cleve, and *Synedrosphenia cuneata* (Grunow) Peragallo (Table 1, Figs. 1-366). The genus with most species and infra-specific representatives was *Mastogloia* with 27 taxa.

DISCUSSION.

Previous floristic studies of benthic diatoms in unexplored areas of the Mexican littorals and the survey of new substrata have yielded numerous new records that have recently enriched the species inventory for the region (Estrada-Gutiérrez *et al.* 2017; López-Fuerte *et al.* 2016; López-Fuerte & Siqueiros Beltrones 2016; Martínez & Siqueiros Beltrones 2018; Siqueiros Beltrones & Argumedo-Hernández 2014; Siqueiros Beltrones & Martínez 2017; Siqueiros Beltrones *et al.* 2017). In this way, the first new diatom taxon for the Gulf of California was recently proposed: *Halamphora primus* López-Fuerte et Siqueiros Beltrones was collected from the skin of stone scorpion fish (López-Fuerte & Siqueiros Beltrones 2018). Thus, the survey of substrates from remote localities such as Islas Revillagigedo

(Mexico) was expected to increase the number of taxa and to demystify their endemic potential. Said expectations are here supported by the new records of known diatom species, as well as the sixteen likely new taxa observed.

Our exploration yielded one of the highest species richness ($S=208$) ever recorded for two samples of epiphytic diatoms. In comparison, a single specimen of *Ploclanium cartilagineum* from the western coast of the Baja California Peninsula, heavily colonized with diatoms, yielded 46 taxa (Siqueiros Beltrones & Argumedo Hernández 2014), while in over eight samples of six rhodophyte taxa from southeastern (tropical) Mexico 115 taxa were identified (Siqueiros Beltrones & Martínez 2017). Another survey still, recorded 143 diatom taxa from ten specimens of *Laurencia pacifica* in five different dates (Siqueiros Beltrones & Hernández-Almeida 2006). Moreover, a much similar study carried out in Isla Guadalupe off Baja California, where transitional oceanographic conditions are characteristic, yielded less (119) taxa (including 11 % new records for Mexican littorals), although in many more samples (López-Fuerte *et al.* 2015). There, *Mastogloia* species, considered mostly of tropical affinity, comprised the higher number of species (13), whilst in the Revillagigedo Islands twice the number of species (27) of this genus were present. Thus, although the Revillagigedo Islands are located also within a transitional zone influenced by the California Current (CONANP-SEMARNAT 2015), the tropical component of the archipelago region does seem to be a major factor enhancing the species diversity of epiphytic diatoms.

Only one taxon, *Grammatophora merletta* Hanna & Grant, was recognized as previously recorded from the Revillagigedo Archipelago by Hanna and Grant (1926) who made a survey on diatoms from the Miocene. On the other hand, most of the identified diatom taxa in Revillagigedo islands have been recorded elsewhere in the Mexican Pacific, particularly in the NW littorals that harbor many temperate forms under transitional oceanographic conditions (López-Fuerte & Siqueiros Beltrones 2016). However, representatives of tropical affinity were also observed, such as the *Mastogloia* specific and infra-specific taxa (27). Plus, other tropical elements that were recorded previously only from the Gulf of Mexico such as *Cocconeis caribensis*, *Mastogloia affirmata*, and *Synedrosphenia cuneata*, and now recorded for the first time in the Mexican Pacific. This agrees with our working hypothesis. However, over 25 % of the taxa are new records hitherto not documented for Mexican littorals (López-Fuerte & Siqueiros Beltrones 2016; Martínez & Siqueiros

Beltrones 2018; Siqueiros Beltrones & Martínez 2017; Siqueiros Beltrones *et al.* 2017) and whose biogeographic affinities are uncertain.

Our results may serve both to demystify the expected endemism and high species diversity of benthic diatoms in the archipelago, inasmuch for the latter the high species richness observed backs said assumption, but most of the recorded taxa are actually distributed worldwide. By contrast, the fact that ten taxa could not be identified after an exhaustive search leads us to infer that these represent new taxa that need formal taxonomic descriptions.

These results, derived from the few samples examined, suggest a likely occurrence of more biogeographically isolated diatom taxa in the islands. As in the case of *Grammatophora monilifera* Tempere & Brun which has been considered solely as a fossil species (Guiry & Gury 2108) from Japan (Kociolek *et al.* 2019). However, although Desikachary (1987) recorded it (single image) in sediments from the Indian Ocean region, it was observed commonly in our samples.

In spite of the low number of hosts surveyed, this first floristic list of epiphytic diatoms of rhodophytes may serve as a reference for upcoming formal and hypothesis driven studies on diatom floristics, taxonomy, ecology and conservation, and biogeography for the Revillagigedo Archipelago. Undertaking the wanting diatom floristics may lead to expect finding over 500 epiphytic taxa on the 190 macroalgae taxa documented (Serviere-Zaragoza *et al.* 2007) in the archipelago, including a significant number of new and, likely, endemic diatom taxa that should require formal taxonomic treatment.

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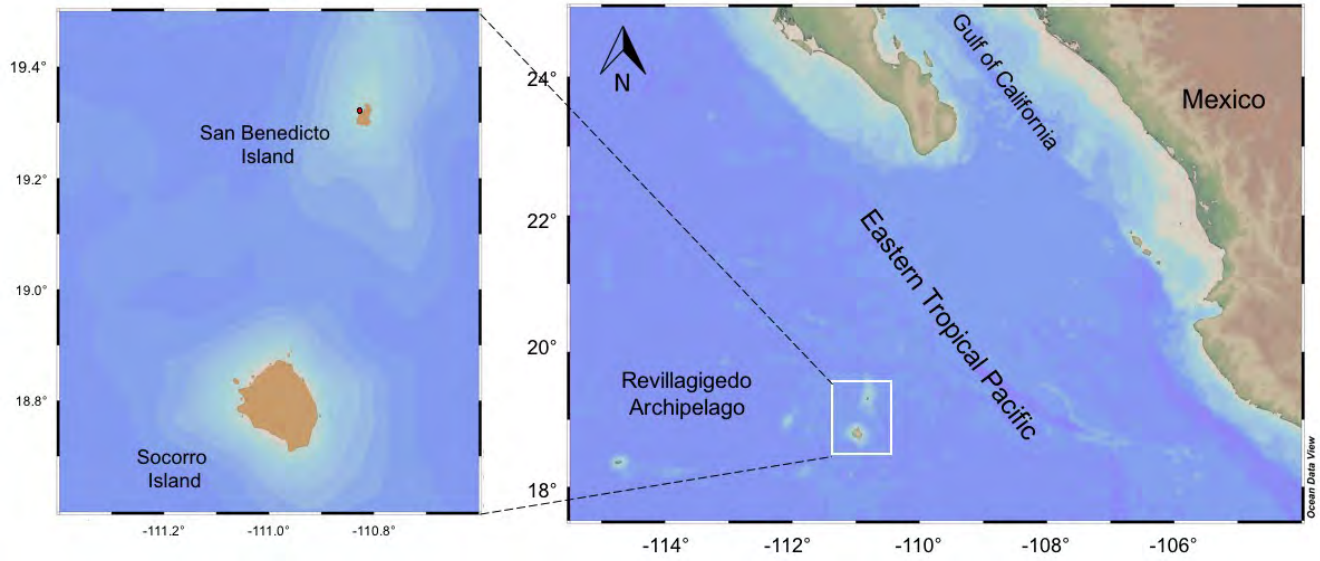
TABLE 1. Floristic list of epiphytic diatoms found on two specimens of *Laurencia* sp. (Rhodophyta) collected at Revillagigedo Islands, Mexico. **NR** = new records for the Mexican coasts; **NRP** = new record for the Mexican Pacific; **NS** = potential new species.

<i>Achnanthes trachyderma</i> (F. Meister) Riaux-Gobin, Compère, Hinz & Ector	NR
<i>Actinocyclus alienus</i> Grunow in Van Heurck	NR
<i>Actinocyclus</i> cf. <i>samoensis</i> (Grunow) De Toni & Forti	NR
<i>Actinocyclus curvatulus</i> Janisch	
<i>Actinocyclus octonarius</i> var. <i>crassus</i> Wm. Smith	
<i>Amphora bigibba</i> Grunow ex A. Schmidt	
<i>Amphora</i> sp. 1 cf. <i>clevei</i> Grunow/ <i>Amphora</i> . cf. <i>eulenstenii</i>	NS
<i>Amphora exilitata</i> Giffen	
<i>Amphora lineolata</i> Ehrenberg	
<i>Amphora maletracta</i> var. <i>constricta</i> (Heiden) Simonsen	
<i>Amphora ostrearia</i> Brébisson ex Kützing	
<i>Amphora proteus</i> Gregory	
<i>Amphora proteus</i> var. <i>contigua</i> Cleve	
<i>Amphora pseudohyalina</i> Simonsen	
<i>Amphora rhombica</i> Kitton	
<i>Amphora staurophora</i> Castracane	
<i>Ardissonea</i> cf. <i>crystallina</i> var. (Agardh) Grunow	NR
<i>Ardissonea formosa</i> (Hantzsch) Grunow	
<i>Ardissonea fulgens</i> (Grevillei) Grunow	
<i>Ardissonea robusta</i> (Ralfs) De Notaris	
<i>Astartiella punctifera</i> Witkowski & Lange-Bertalot	
<i>Asteromphalus petersoni</i> (Kolbe) Thorrington-Smith	
<i>Bleakeleya notata</i> (Grunow) Round	
<i>Caloneis elongata</i> (liber) (Grunow) Cleve	
<i>Caloneis latiuscula</i> f. <i>minor</i> Foged	
<i>Caloneis maxima</i> var. <i>excentrica</i> Grunow	NR

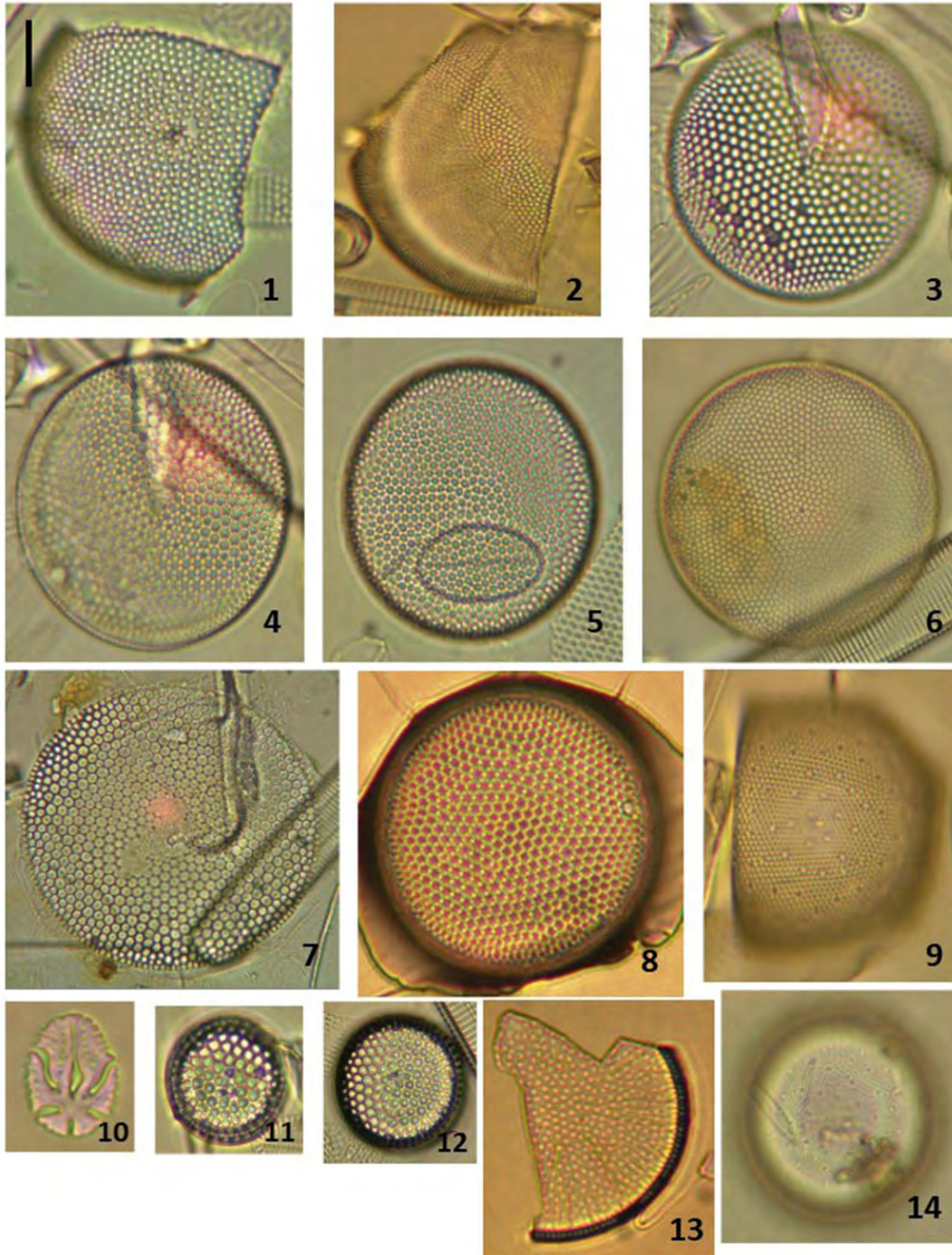
<i>Caloneis linearis</i> (Grunow) Boyer	
<i>Caloneis</i> sp. 1	NS
<i>Caloneis</i> sp. 2 cf. <i>linearis</i>	NR
<i>Caloneis</i> sp. 3	NS
<i>Caloneis</i> sp. 4	NS
<i>Caloneis</i> sp. 5 (“cymbelliformis”, Encyonema like)	NS
<i>Campylodiscus neofastuosus</i> Ruck & Nakov	
<i>Campylodiscus thuretii</i> Brébisson	
<i>Campylodiscus decorus</i> Brébisson	
<i>Catenula</i> sp.	NR
<i>Climacosphenia moniligera</i> Ehrenberg	
<i>Cocconeis caribensis</i> Romero & Navarro	NRP
<i>Cocconeis comis</i> A.W.F. Schmidt	NR
<i>Cocconies composita</i> A. Schmidt	NR
<i>Cocconeis convexa</i> Giffen	
<i>Cocconeis diminuta</i> (Pantocsek) Hustedt	
<i>Cocconeis dirupta</i> Gregory	
<i>Cocconeis dirupta</i> var. <i>flexella</i> (Janisch & Rabenhorst) Grunow	
<i>Cocconeis diruptoides</i> Hustedt	
<i>Cocconeis heteroidea</i> Hantzsch	
<i>Cocconeis krammeri</i> Lange-Bertalot & Metzeltin	
<i>Cocconeis molesta</i> Kützing	
<i>Cocconeis molesta</i> var. <i>molesta</i> Witkowski, Lange-Bertalot & Metzeltin	NR
<i>Cocconeis placentula</i> Ehrenberg	
<i>Cocconeis placentula</i> var. <i>euglypta</i> (Ehrenberg) Cleve	
<i>Cocconeis pseudodiruptoides</i> Foged	
<i>Cocconeis scutellum</i> Ehrenberg	
<i>Cocconeis scutellum</i> var. <i>parva</i> Grunow	
<i>Cocconeis stauroneiformis</i> (Van Heurck) Okuno	
<i>Cocconeis vetusta</i> A. Schmidt	
<i>Cocconeis</i> sp. 1	
<i>Cocconeis</i> sp. 2	NS
<i>Cocconeis</i> sp. 3	NS
<i>Coscinodiscus radiatus</i> Ehrenberg	
<i>Cyclophora tenuis</i> Castracane var. <i>tropica</i> Grunow	NR
<i>Cymbellonitzschia banzuensis</i> Stepanek, Hamsher, Mayama, Jewson & Kociolek	NR
<i>Cymbellonitzschia</i> sp. 2	NR
<i>Delphineis minutissima</i> (Hustedt) Simonsen	
<i>Dimerogramma minor</i> var. <i>nana</i> (Gregory) Van Heurck	
<i>Diploneis chersonensis</i> Cleve	
<i>Diploneis crabro</i> Ehrenberg	
<i>Diploneis crabro</i> f. <i>dirhombus</i> (A. Schmidt) Hustedt	NR
<i>Diploneis litoralis</i> (Donkin) Cleve	
<i>Diploneis litoralis</i> var. <i>clathrata</i> Østrup	
<i>Diploneis parca</i> (A. Schmidt) Boyer	NR
<i>Diploneis</i> cf. <i>peterseni</i> Hustedt	
<i>Diploneis smithii</i> (Brébisson) Cleve	
<i>Diploneis vacillans</i> var. <i>renitens</i> A. Schmidt	
<i>Diploneis vacillans</i> var. <i>vacillans</i> Witkowski, Lange-Bertalot & Metzeltin	
<i>Ehrenbergiulva granulosa</i> (Grunow) Witkowski, Lange-Bertalot & Metzeltin	
<i>Epithemia pacifica</i> (Krammer) Lobban & Park	
<i>Falcula media</i> Voigt	NR
<i>Fallacia inscriptura</i> (Hendey) Witkowski, Lange-Bertalot & Metzeltin	
<i>Frustulia interposita</i> (Lewis) De Toni	
<i>Gomphonemopsis pseudexigua</i> (Simonsen) Medlin	
<i>Grammatophora marina</i> (Lyngbye) Kützing	
<i>Grammatophora macilenta</i> W. Smith	
<i>Grammatophora merletta</i> Hanna & Grant	NR
<i>Grammatophora oceanica</i> (Ehrenberg pro parte) Grunow	
<i>Grammatophora oceanica</i> var. <i>nodulosa</i> Grunow	NR
<i>Grammatophora undulata</i> Ehrenberg	
<i>Grammatophora monilifera</i> Tempere & Brun	NS
<i>Grammatophora undulata</i> var. <i>gallopagensis</i> Grunow	NR
<i>Gyrosigma tenuissimum</i> (W. Smith) Griffith & Henfrey	
<i>Halamphora angularis</i> (Gregory) Levkov	

<i>Halamphora</i> cf. <i>staurophora</i> (Juhlin-Dannfelt) Álvarez-Blanco & Blanco	NR
<i>Halamphora coffeaeformis</i> (C. Agardh) Levkov	
<i>Halamphora dusenii</i> (Brun) Levkov	
<i>Halamphora staurophora</i> (Juhlin-Dannfelt) Álvarez-Blanco & Blanco	
<i>Halamphora turgida</i> (Gregory) Levkov	
<i>Hantzschia amphioxys</i> (Ehrenberg) Grunow	
<i>Haslea</i> cf. <i>howeana</i> (Hagelstein) Giffen	
<i>Haslea nautica</i> (Cholnoky) Giffen	NR
<i>Hyalosynedra laevigata</i> (Grunow) Williams & Round	
<i>Hyalosira tropicalis</i> Navarro	
<i>Licmophora abbreviata</i> Agardh	
<i>Licmophora debilis</i> (Kützing) Grunow in Van Heurck	
<i>Licmophora ehrenbergii</i> (Kützing) Grunow	
<i>Licmophora flabellata</i> (Carmichael) Agardh	
<i>Licmophora gracilis</i> (Ehrenberg) Grunow	
<i>Licmophora paradoxa</i> (Lyngbye) Agardh	
<i>Licmophora remulus</i> Grunow	
<i>Lioloma delicatulum</i> Hasle	NR
<i>Lyrella clavata</i> var. <i>caribaea</i> (Cleve) Siqueiros-Beltrones	
<i>Mastogloia acutiuscula</i> var. <i>elliptica</i> Hustedt	
<i>Mastogloia affinis</i> Cleve	
<i>Mastogloia affirmata</i> (Leudiger-Fortmore) Cleve	NRP
<i>Mastogloia apiculata</i> Wm. Smith	
<i>Mastogloia binotata</i> (Grunow) Cleve	
<i>Mastogloia borneensis</i> Hustedt	
<i>Mastogloia ciskeiensis</i> Giffen	
<i>Mastogloia cocconeiformis</i> Grunow	NR
<i>Mastogloia corsicana</i> Grunow in Cleve & Möller	
<i>Mastogloia crucicula</i> (Grunow) Cleve	
<i>Mastogloia crucicula</i> var. <i>alternans</i> Zanoni, Novarino & Bazzichelli	
<i>Mastogloia cuneata</i> (Meister) Simonsen	
<i>Mastogloia delicatissima</i> Hustedt	NR
<i>Mastogloia emarginata</i> Hustedt	
<i>Mastogloia erythraea</i> Grunow	
<i>Mastogloia exilis</i> Hustedt	
<i>Mastogloia fimbriata</i> (Brightwell) Cleve	
<i>Mastogloia grunowii</i> A. Schmidt	
<i>Mastogloia horvathiana</i> Grunow	
<i>Mastogloia inaequalis</i> Cleve	
<i>Mastogloia marginulata</i> Grunow	NR
<i>Mastogloia ovulum</i> Hustedt	
<i>Mastogloia ovum-paschale</i> (A. Schmidt) Mann	
<i>Mastogloia parva</i> Hustedt	NR
<i>Mastogloia punctatissima</i> (Greville) Ricard	
<i>Mastogloia pusilla</i> var. <i>subcapitata</i> Hustedt	
<i>Mastogloia quinquecostata</i> Grunow	NR
<i>Mastogloia subaffirmata</i> Hustedt	
<i>Melosira montagnei</i> (Kützing) Lagersted	
<i>Microtabella</i> sp.	NS
<i>Navicula abunda</i> Hustedt	
<i>Navicula cincta</i> (Ehrenberg) Ralfs	
<i>Navicula comoides</i> (Dillwyn) H. Peragallo	
<i>Navicula halophila</i> (Grunow) Cleve	
<i>Navicula johanrossii</i> Giffen	
<i>Navicula leptoloba</i> Meister	NR
<i>Navicula libellus</i> Gregory	
<i>Navicula longa</i> (Gregory) Ralfs	
<i>Navicula zostereti</i> Grunow	
<i>Neosynedra provincialis</i> (Grunow) Williams & Round	
<i>Nitzschia bicapitata</i> Cleve	
<i>Nitzschia bombiformis</i> Grunow	NR
<i>Nitzschia confinis</i> Hustedt	
<i>Nitzschia dissipata</i> (Kützing) Grunow	
<i>Nitzschia distans</i> Gregory	
<i>Nitzschia lanceolata</i> W. Smith	

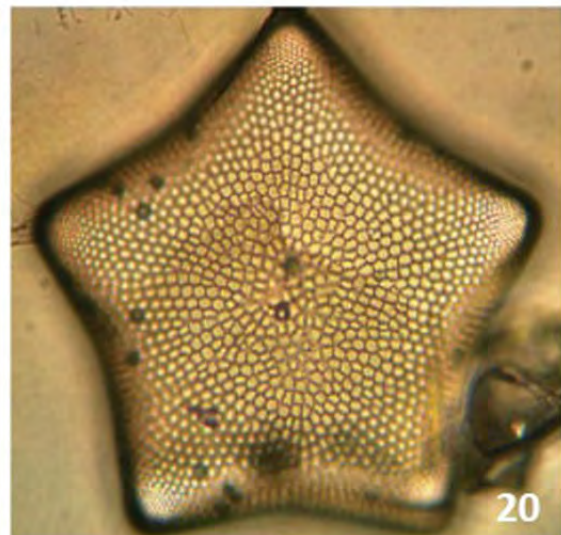
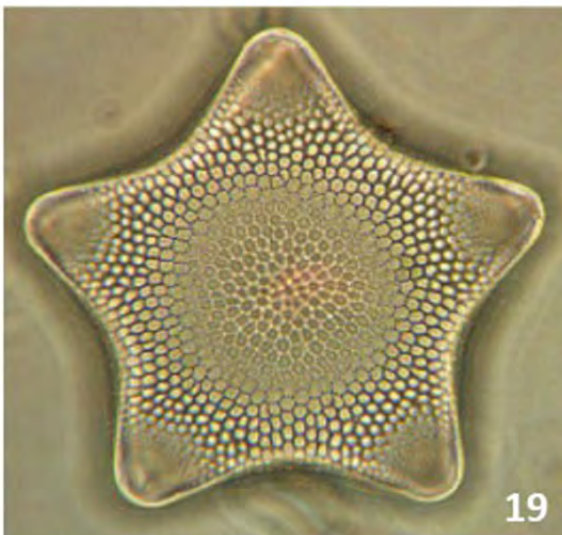
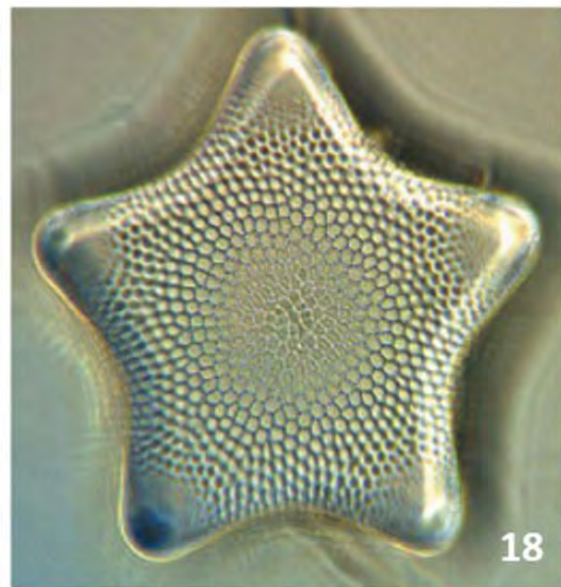
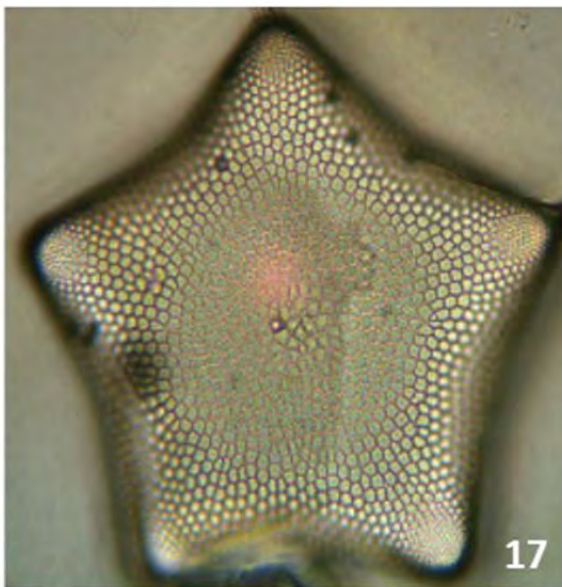
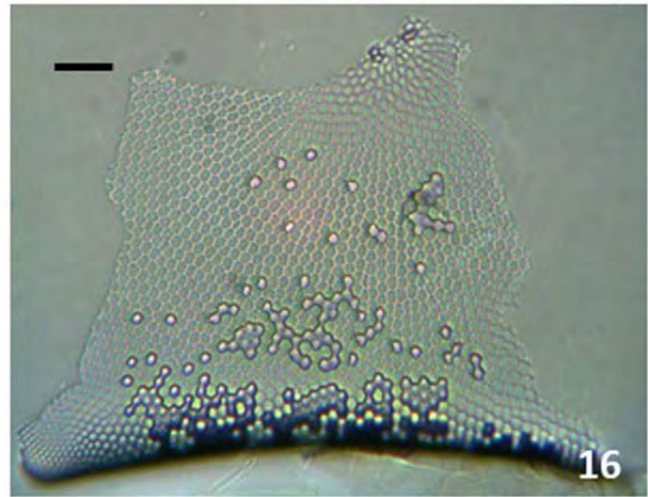
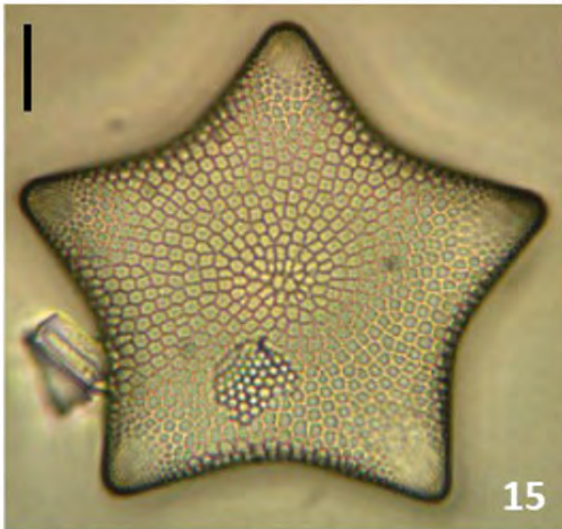
<i>Nitzschia lanceolata</i> var. <i>minor</i> (Grunow) H. Peragallo & M. Peragallo	
<i>Nitzschia longissima</i> f. <i>costata</i> Hustedt	
<i>Nitzschia microcephala</i> var. <i>bicapitellata</i> A. Cleve	
<i>Nitzschia pellucida</i> Grunow	
<i>Nitzschia persuadens</i> Cholnoky	
<i>Nitzschia</i> cf. <i>panduriformis</i> var. <i>continua</i> Grunow	
<i>Nitzschia sicula</i> (Castracane) Hustedt	
<i>Nitzschia sigma</i> (Kützing) W. Smith	
<i>Nitzschia sigma</i> (Kützing) W. Smith var.	
<i>Nitzschia subacuta</i> Hustedt	NR
<i>Nitzschia tubicola</i> Grunow	NR
<i>Odontidium</i> sp.	NS
<i>Neofragilaria anomala</i> Witkowski & Dabek	NR
<i>Parlibellus cruciculoides</i> (Brockman) Witkowski, Lange-Bertalot & Metzeltin	
<i>Parlibellus</i> cf. <i>phoebeae</i> Witkowski, Metzeltin & Lange-Bertalot	NR
<i>Plagiodiscus nervatus</i> Grunow	
<i>Pleurosigma distinguendum</i> Hustedt	NR
<i>Podocystis americana</i> Bailey	
<i>Pravifusus</i> sp.	NS
<i>Psammodictyon constrictum</i> (Kützing) D.G. Mann	
<i>Psammodictyon constrictum</i> var.	
<i>Pseudohimantidium</i> sp.	NS
<i>Pteroncola</i> sp.	NS
<i>Rhabdonema adriaticum</i> Kützing	
<i>Rhoicosphenia</i> cf. <i>genuflexa</i> (Kützing) Medlin	
<i>Rhopalodia gibberula</i> (Ehrenberg) O. Müller	
<i>Rhopalodia gibberula</i> var. <i>producta</i> (Grunow) O. Müller	
<i>Rhopalodia</i> sp. (epithemioide)	NS
<i>Roperia tessellata</i> (Roper) Grunow	
<i>Seminavis delicatula</i> Wachnicka & Gaiser	
<i>Seminavis</i> sp. (Amphora sp.?)	
<i>Seminavis strigosa</i> (Hustedt) Danieledis & Economou-Amilli	
<i>Shionodiscus oestrupii</i> (Ostenfeld) Alverson, Kang & Theriot	
<i>Striatella</i> cf. <i>interrupta</i> (Ehrenberg) Heiberg	
<i>Striatella delicatula</i> Kützing	
<i>Synedra commutata</i> Grunow	
<i>Synedrosphenia cuneata</i> (Grunow) Peragallo	NRP
<i>Tabularia barbatula</i> (Kützing) Williams & Round	
<i>Tabularia fasciculata</i> (C. Agardh) Williams & Round	
<i>Tabularia investiens</i> (W. Smith) Williams & Round	
<i>Thalassionema nitzschioides</i> (Grunow) Van Heurck	
<i>Toxarium hennedyanum</i> (Gregory) Pelletan	
<i>Toxarium undulatum</i> Bailey	
<i>Trachyneis aspera</i> (Ehrenberg) Cleve	
<i>Trachyneis aspera</i> var. <i>oblonga</i> (Bailey) Cleve	NR
<i>Triceratium formosum</i> f. <i>quiquelobatum</i> (Greville) Hustedt	NR
<i>Trigonium formosum</i> (Brightwell) Hendey	
<i>Tropidoneis</i> sp.	NS
<i>Tropidoneis pusilla</i> (W.Gregory) Cleve	
<i>Tropidoneis vitrea</i> (W.M.Smith) Cleve & Möller	
<i>Tryblionella coarctata</i> (Grunow) D.G. Mann	
<i>Tryblionella marginulata</i> var. <i>didyma</i> (Grunow) Haworth & Kelly	



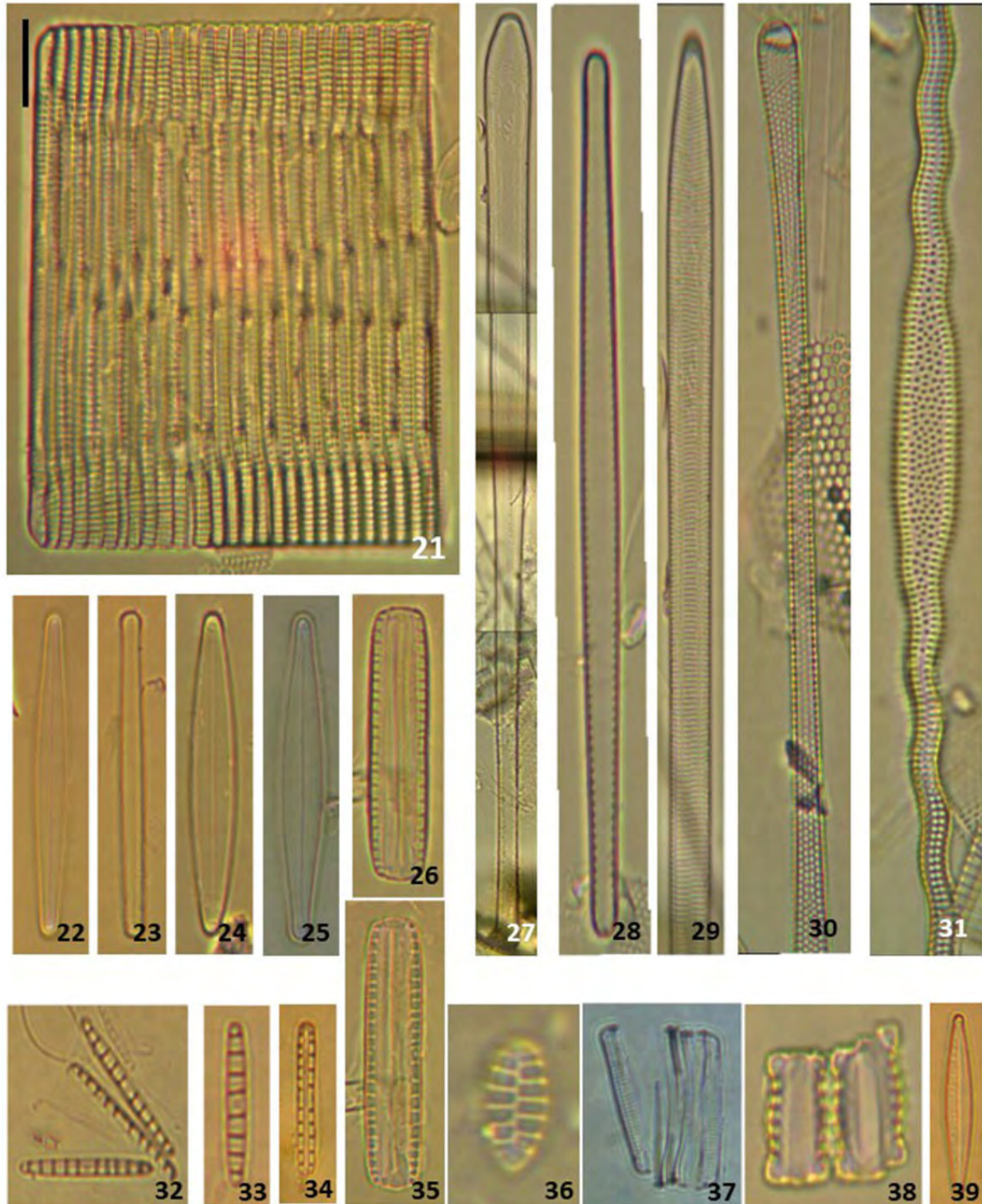
Map 1. Location of the Revillagigedo Archipelago and sampling site at San Benedicto island



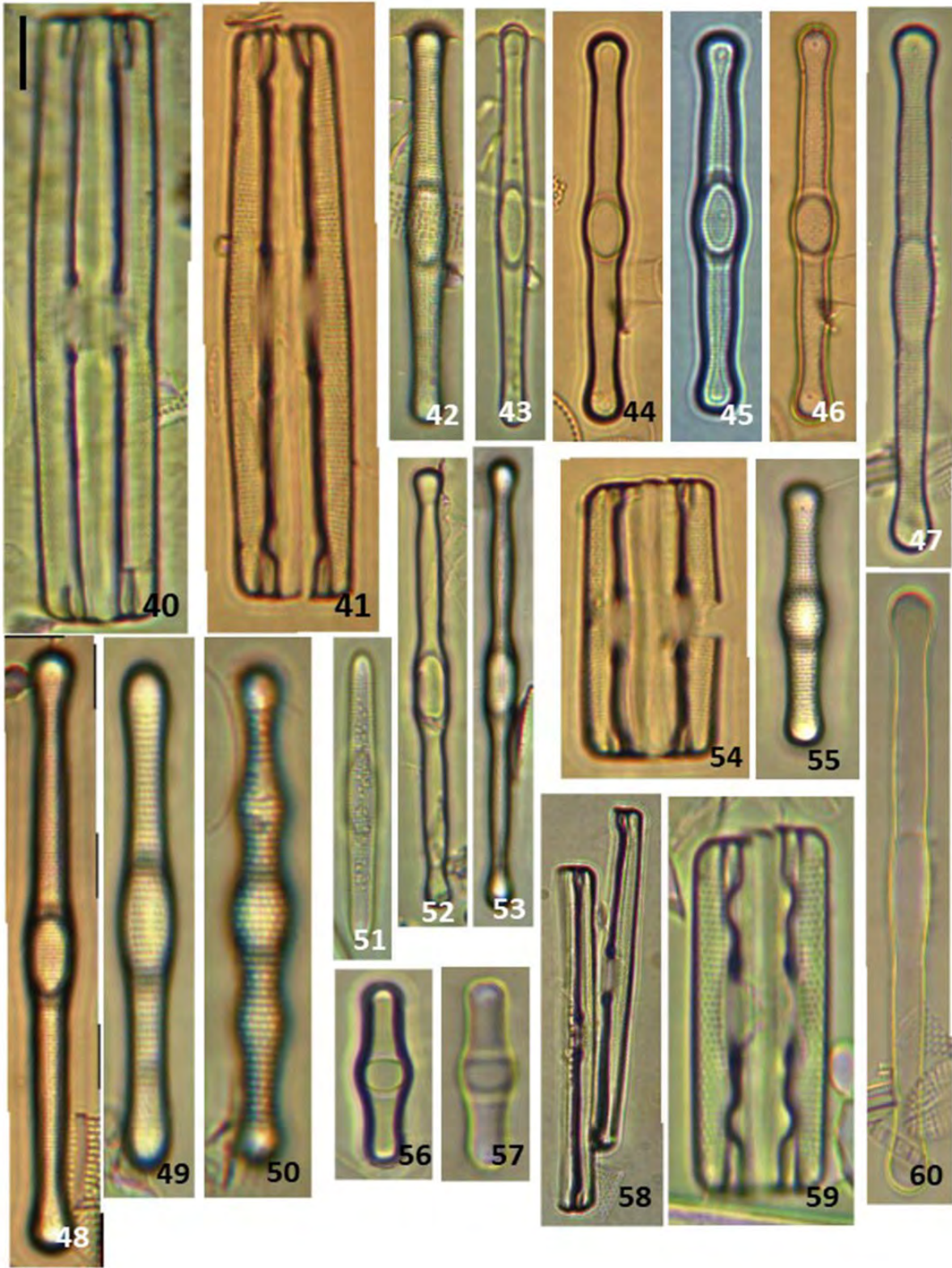
Figures 1- 14. 1. *Actinocyclus octonarius* var. *crassus*; 2. *Actinocyclus* cf. *samoensis*; 3, 4. *Actinocyclus curvatulus*; 5, 7. *Coscinodiscus radiatus*; 6. *Actinocyclus alienus*; 8. *Roperia tessellata*; 9, 14. *Melosira montagnei*; 10. *Asteromphalus petersonii*; 11, 12. *Shionodiscus oestrupii*; 13-14. *Ehrenbergiulva granulosa*.
 En esta lámina y en las siguientes la barra equivale a 10 micrómetros.



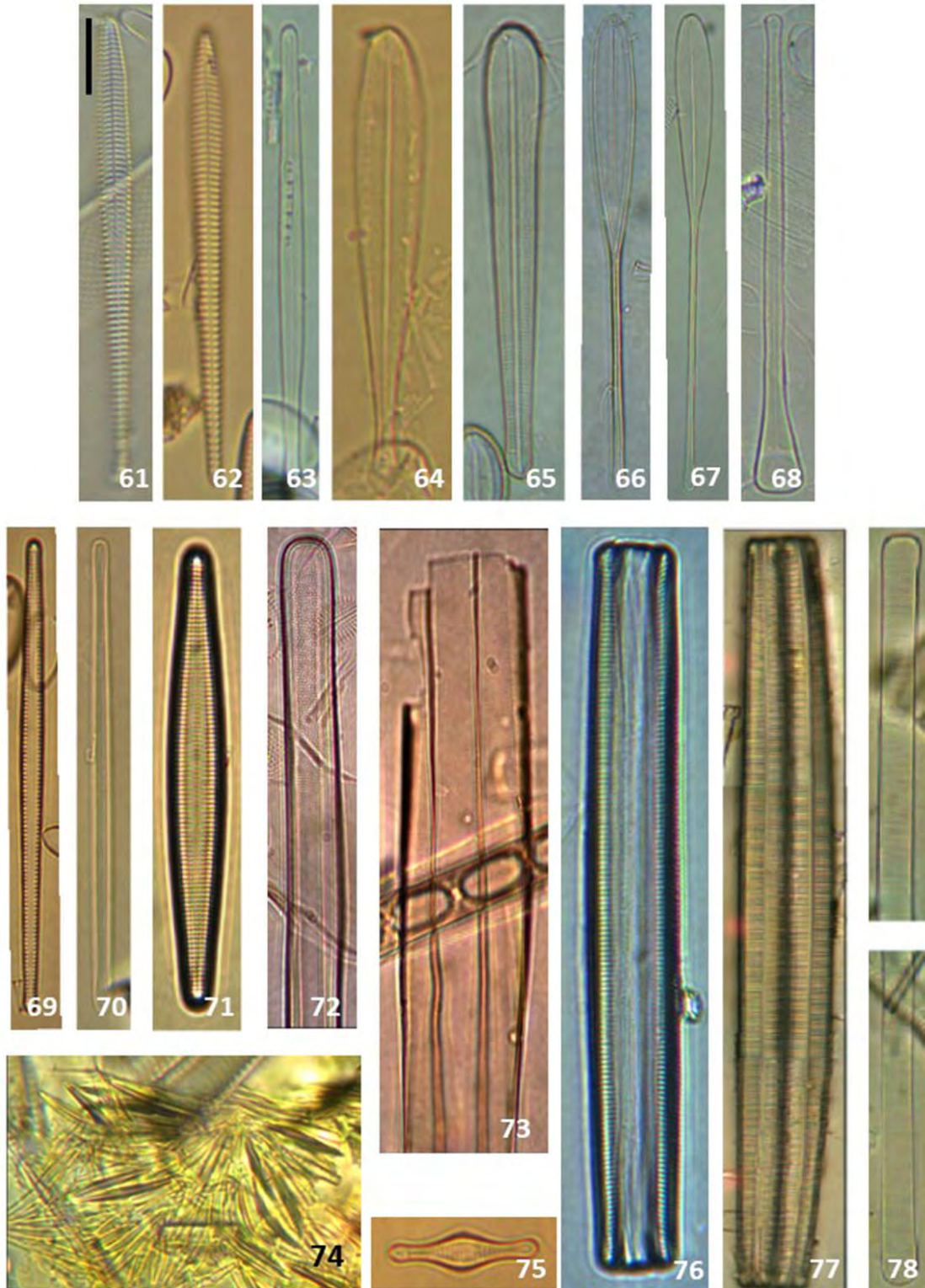
Figures 15-20. 15, 17-20. *Triceratium formosum* f. *quinguelobatum*; 16. *Trigonium formosum*.



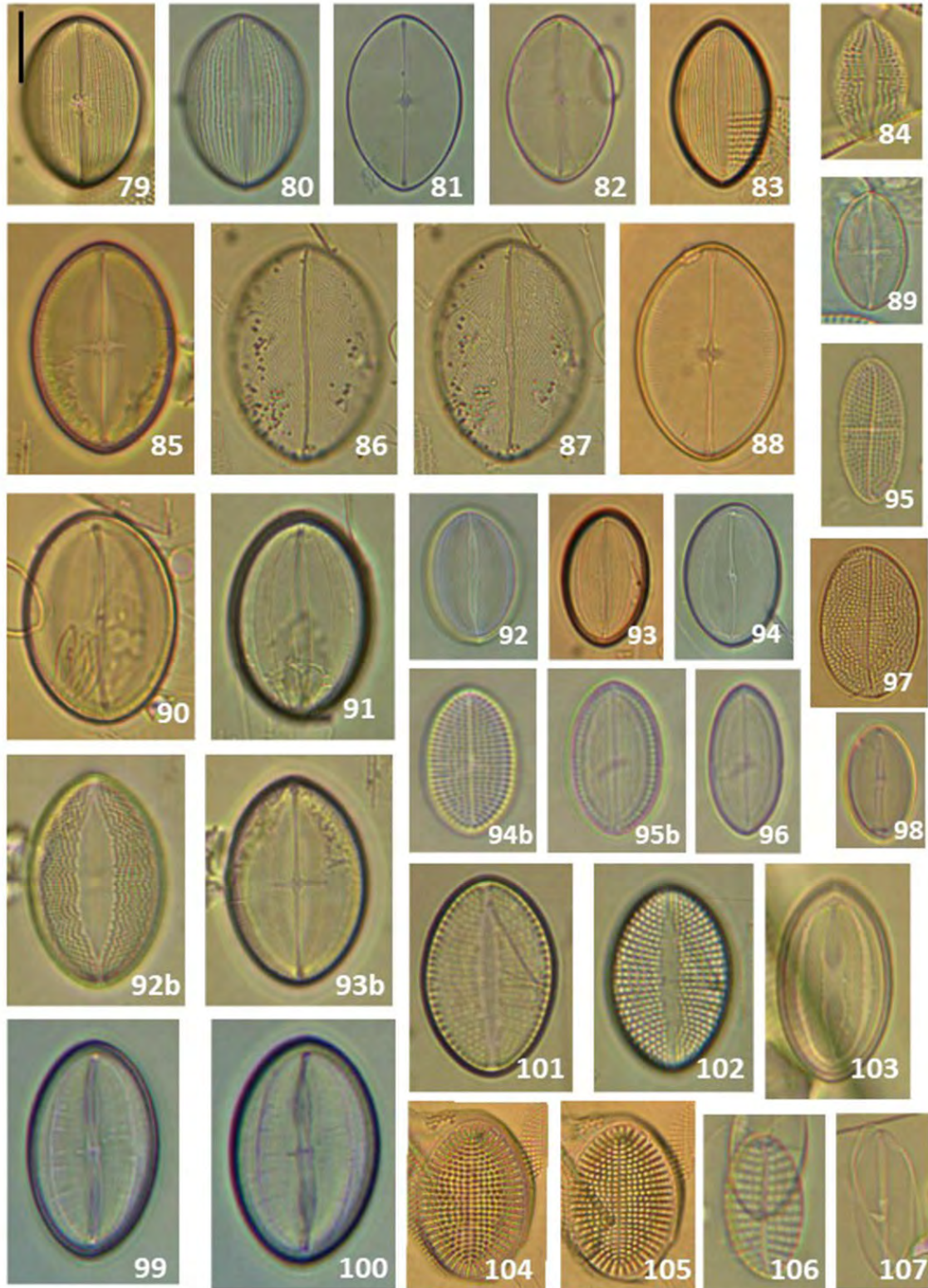
Figures 21-39. 21. *Rhabdonema adriaticum*; 22, 25. *Hyalosynedra laevigata*; 23. *Neosynedra provincialis*; 24. *Synedra commutata*; 26, 32-35. *Odontidium* sp.; 27, 29. *Synedrosphenia cuneata*; 28. *Thalassionema nitzschioides*; 30. *Toxarium hennedyanum*; 31. *Toxarium undulatum*; 36. *Neofragilaria anomala*; 37. *Striatella delicatula*; 38. *Dimerogramma minor* var. *nana*; 39. *Tabularia barbatula*.



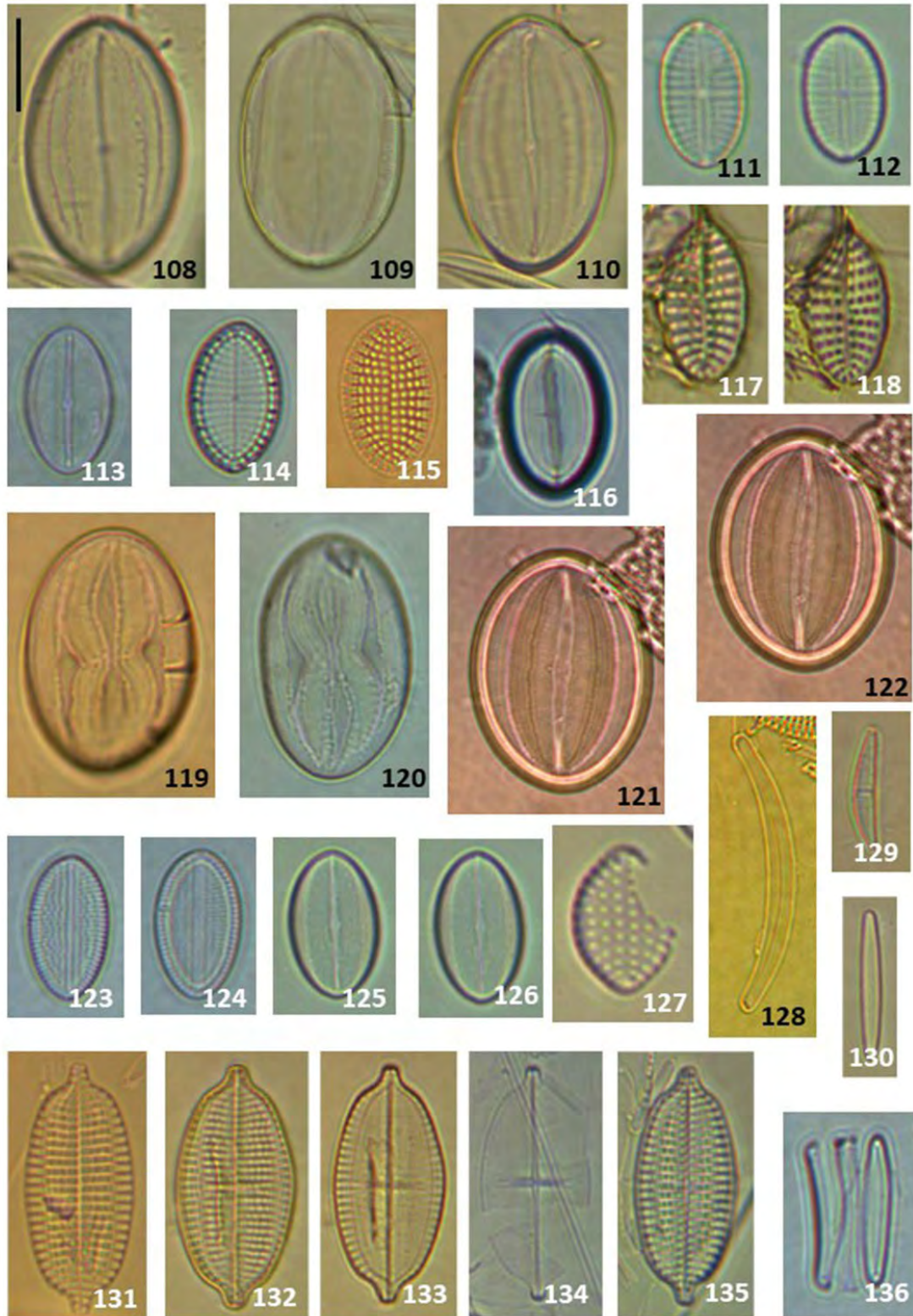
Figures 40-60. 40, 51. *Grammatophora oceanica*; 41-43. *Grammatophora macilentia*; 44-47, 60. *Grammatophora merletta*; 48, 49, 55, 58. *Grammatophora monilifera*; 50. *Grammatophora undulata*; 52, 53. *Grammatophora undulata* var. *gallopagensis*; 54, 59. *Grammatophora marina*; 56, 57. *Grammatophora oceanica* var. *nodulosa*.



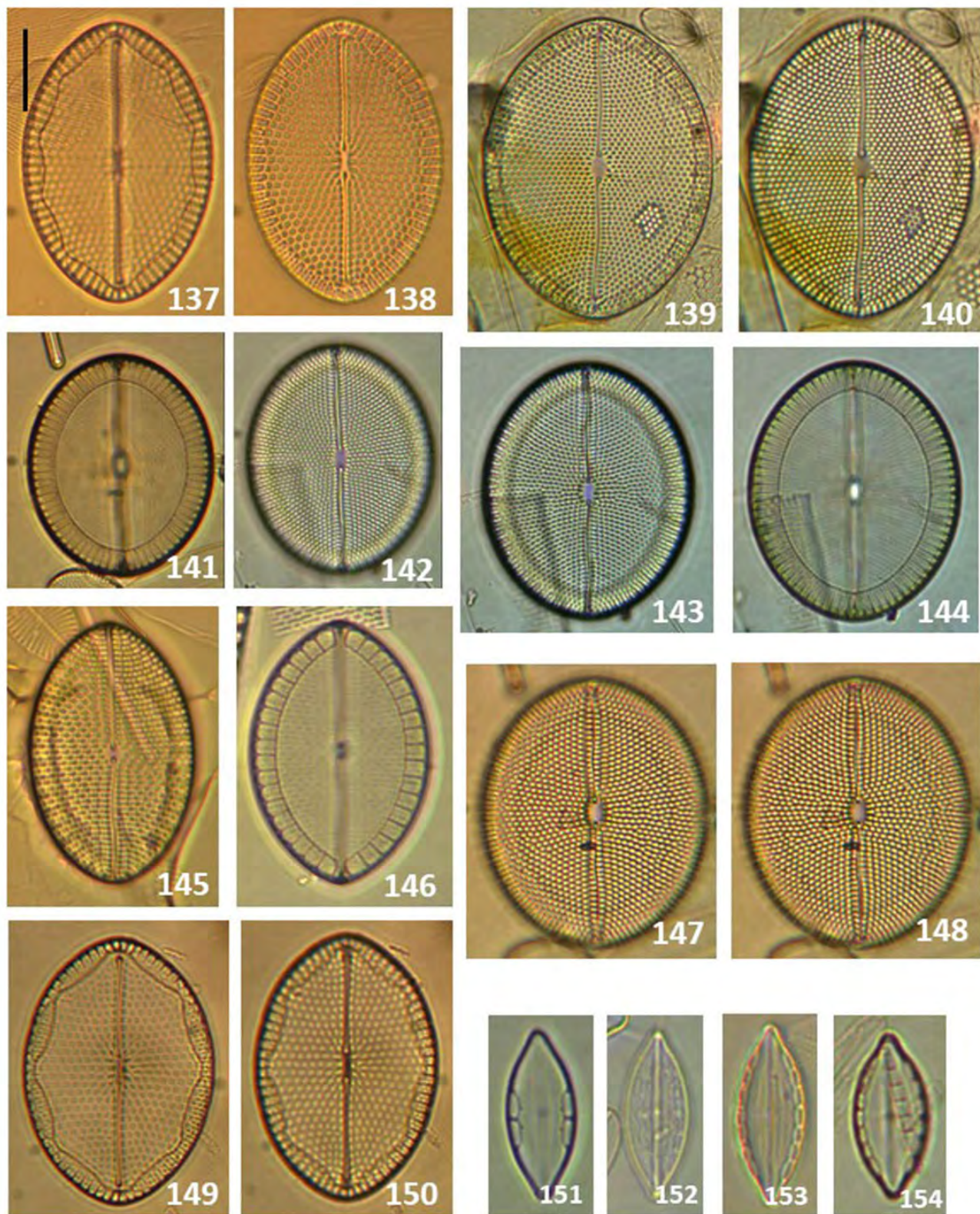
Figures 61-78. 61, 62. *Licmophora abbreviata*; 63. *Hyalosynedra laevigata*; 64. *Licmophora gracilis*; 65. *Licmophora paradoxa*; 66, 67. *Licmophora remulus*; 68. *Bleakeleya notata*; 69. *Tabularia fasciculata*; 70. *Lioloma delicatulum*; 71. *Synedra commutata*; 72, 73. *Climacosphenia moniligera*; 74. *Pseudohimantidium* sp.; 75. *Hyalosira tropicalis*; 76. *Ardissonea formosa*; 77. *Ardissonea robusta*; 78. *Ardissonea fulgens*.



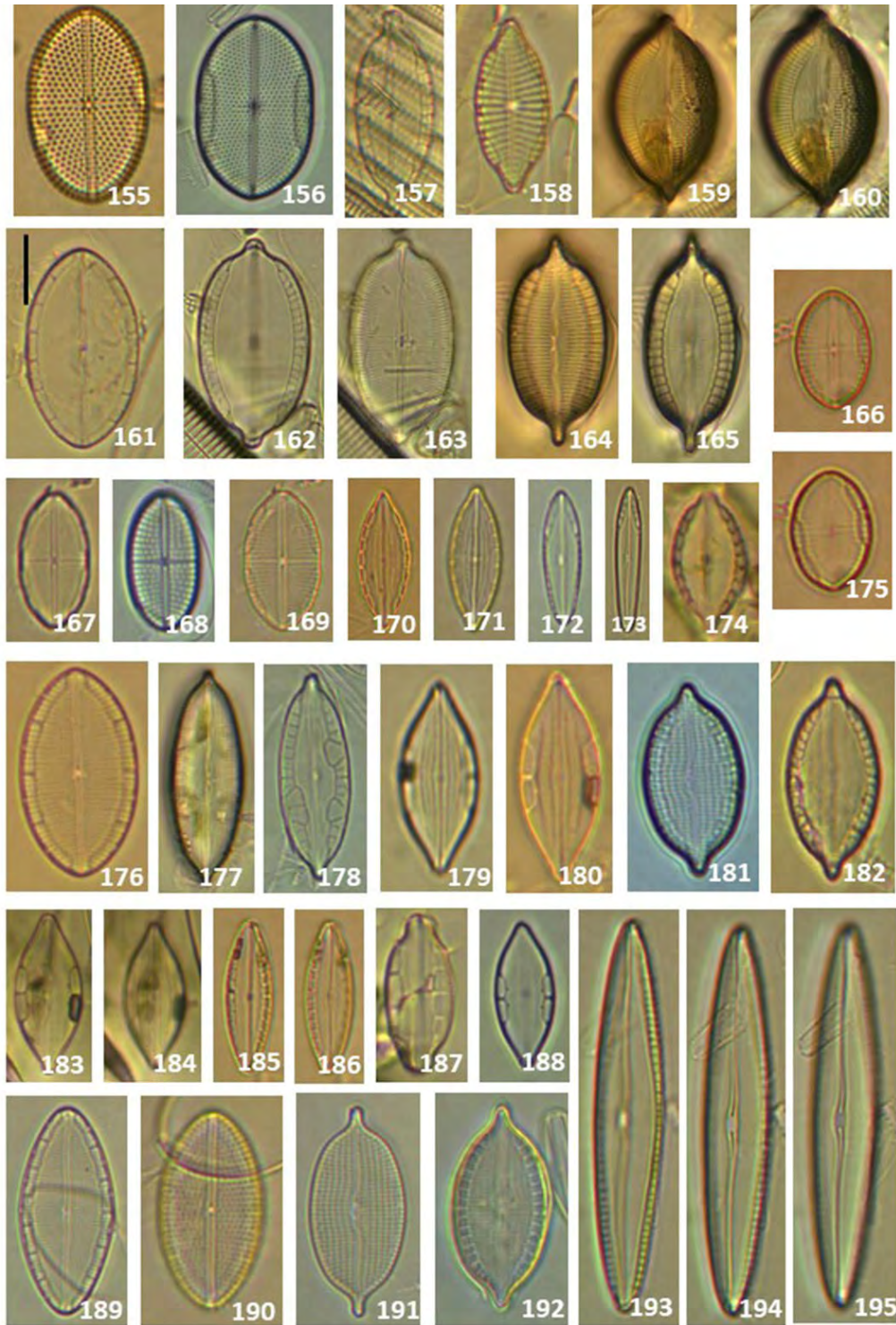
Figures 79-107. 79-83, 88. *Cocconeis convexa*; 84, 89. *Cocconeis dirupta* var. *flexella*; 85, 92, 93. *Cocconeis vetusta*; 86, 87, 90. *Cocconeis dirupta*; 91, 94. *Cocconeis heteroidea*; 92b, 93b, 99, 100. *Cocconeis krammeri*; 94b, 95b, 96. *Cocconeis placentula*; 95. *Cocconeis pseudodiruptoides*; 97. *Cocconeis diruptoides*; 98. *Cocconeis* sp. 2; 101, 102. *Cocconeis* sp. 3; 103. *Cocconeis comis*; 104, 105. *Cocconeis scutellum*; 106. *Cocconeis stauroneiformis*; 107. *Astartiella punctifera*.



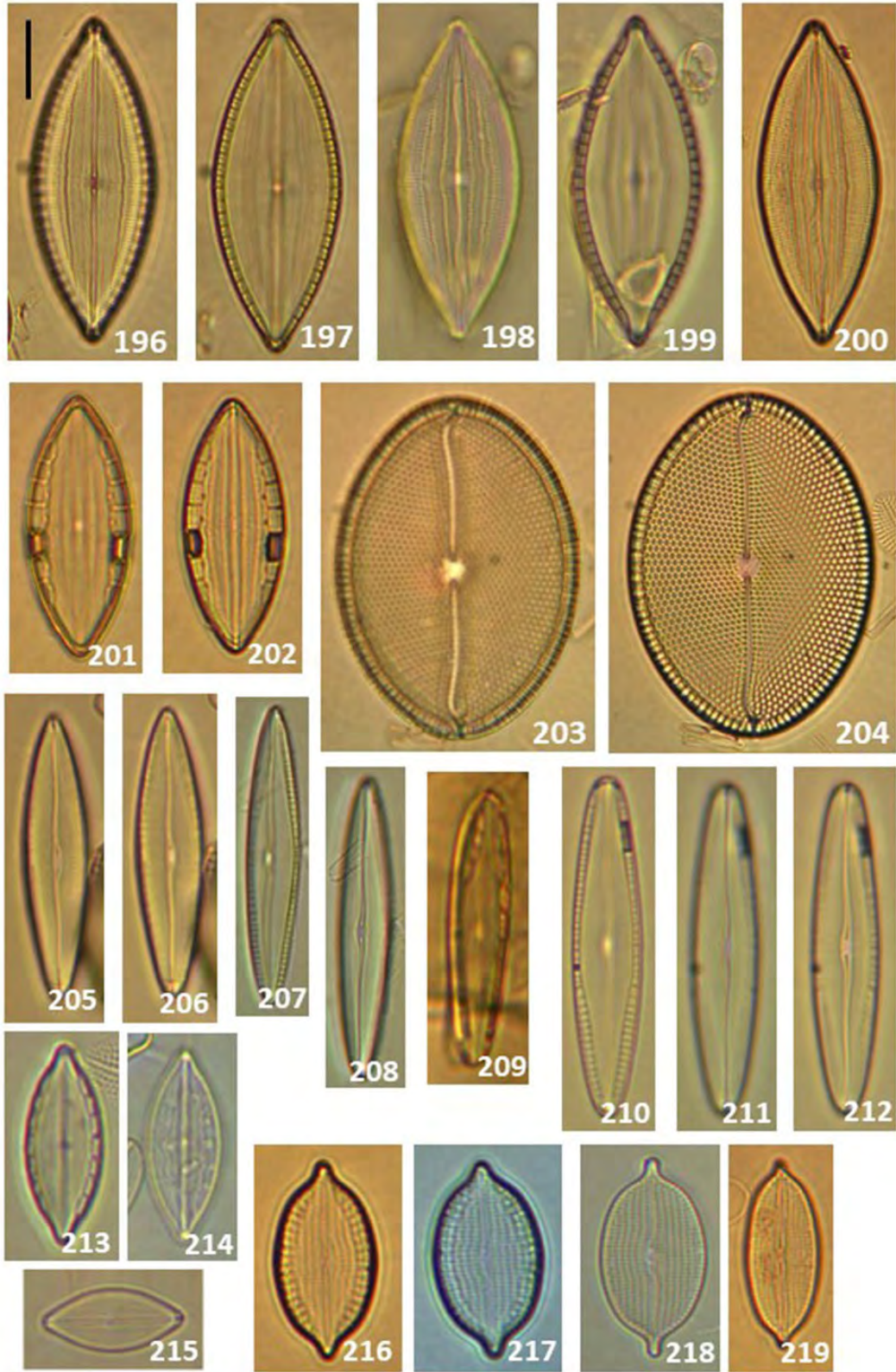
Figures 108-136. 108-110, 121, 122. *Cocconeis heteroidea*; 111, 112. *Mastogloia crucicula*; 113, 125, 126. *Cocconeis* sp. 1; 114, 115. *Cocconeis scutellum* var. *parva*; 116. *Cocconeis krammeri*; 117, 118. *Cocconeis diminuta*; 119, 120. *Cocconeis caribensis*; 123, 124. *Cocconeis placentula* var. *euglypta*; 127. *Delphineis minutissima*; 128. *Falcula media*; 129. *Amphora staurophora*; 130, 136. *Pravifusus* sp.; 131-135. *Achnanthes trachyderma*.



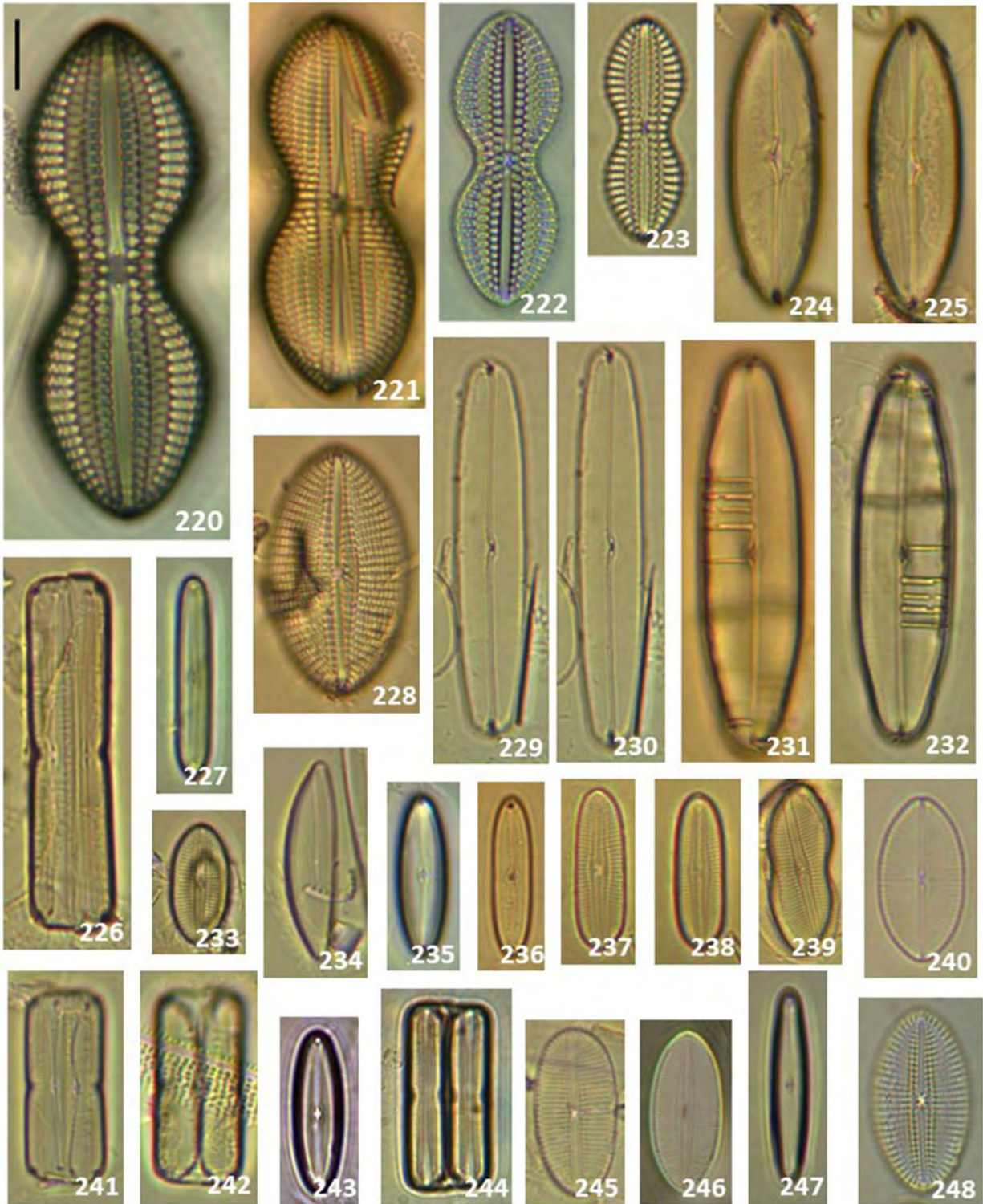
Figures 137-154. 137, 138, 149, 150. *Mastogloia fimbriata*; 139, 140. *Mastogloia horvathiana*; 141-144, 147, 148. *Mastogloia cocconeiformis*; 145, 146. *Mastogloia ovum-paschale*; 151. *Mastogloia exilis*; 152-154. *Mastogloia borneensis*.



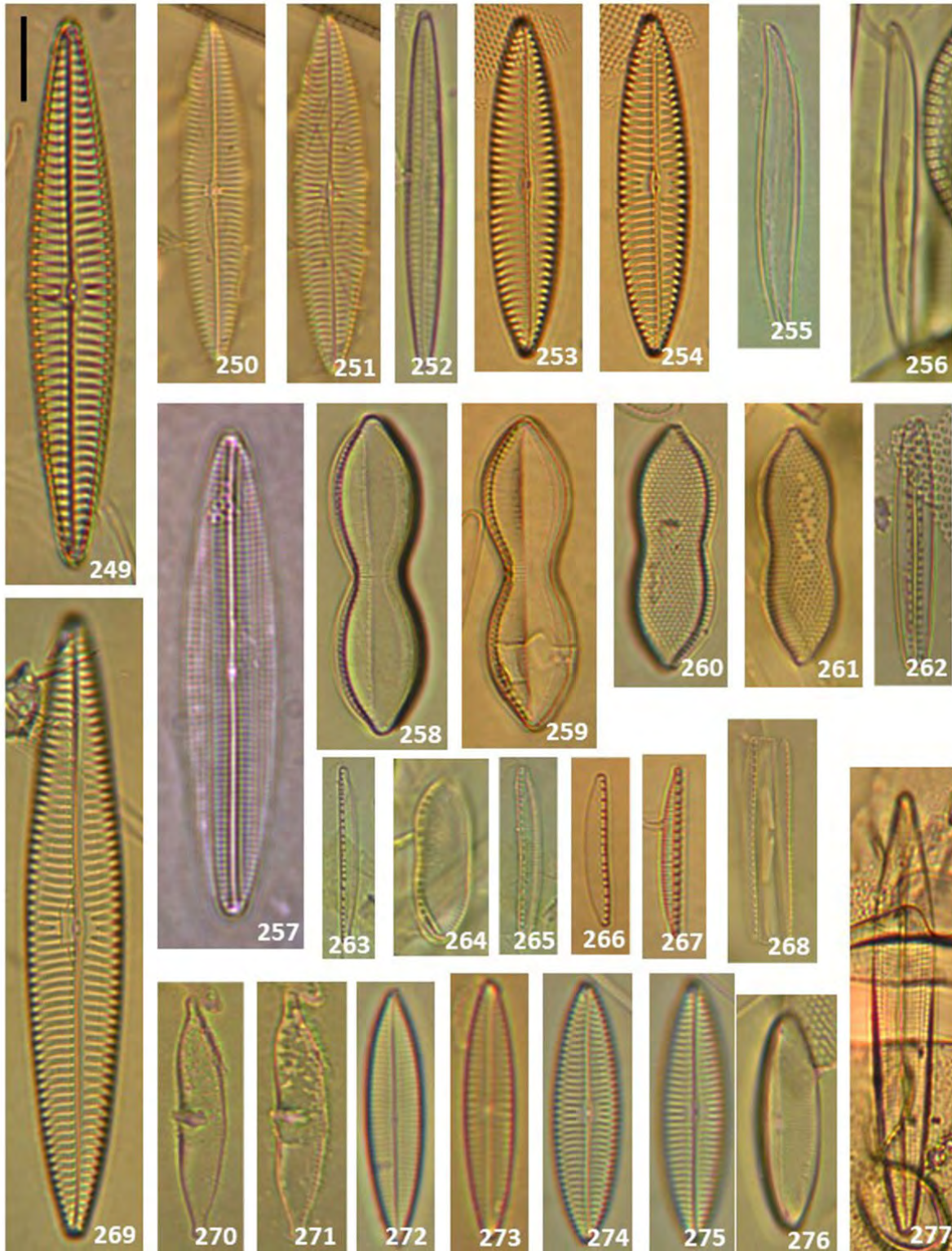
Figures 155-195. 155, 156. *Mastogloia binotata*; 157. *Mastogloia acutiuscula* var. *elliptica*; 158. *Mastogloia affinis*; 159, 160. *Mastogloia affirmata*; 161, 189, 190. *Mastogloia ovulum*; 162, 163. *Mastogloia apiculata*; 164, 165. *Mastogloia corsicana*; 166, 175. *Mastogloia crucicula* var. *alternans*; 167-169. *Mastogloia crucicula*; 170, 171. *Mastogloia borneensis*; 172, 173. *Mastogloia cuneata*; 174. *Mastogloia delicatissima*; 176. *Mastogloia emarginata*; 177, 178. *Mastogloia erythraea*; 179, 180, 183, 184, 188. *Mastogloia exilis*; 181, 182, 191, 192. *Mastogloia subaffirmata*; 185, 186. *Mastogloia marginulata*; 187. *Mastogloia pusilla* var. *subcapitata*; 193-195. *Mastogloia ciskeiensis*.



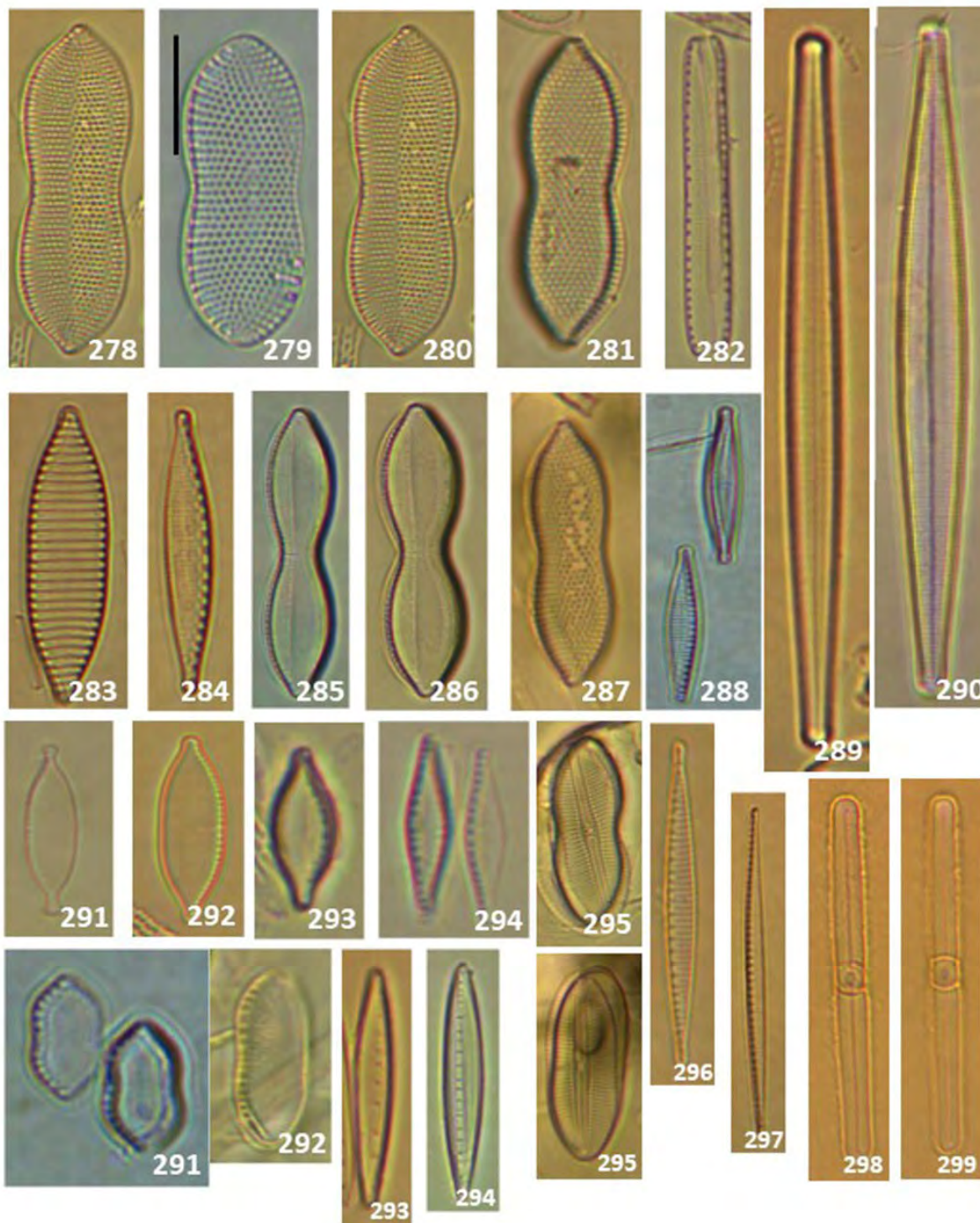
Figures 196-219. 196-200. *Mastogloia quinquecostata*; 201, 202. *Mastogloia grunowii*; 203, 204. *Mastogloia punctatissima*; 205-208. *Mastogloia ciskeiensis*; 209. *Mastogloia cuneata*; 210-212. *Mastogloia inaequalis*; 213-215. *Mastogloia borneensis*; 216-218. *Mastogloia subaffirmata*; 219. *Mastogloia parva*.



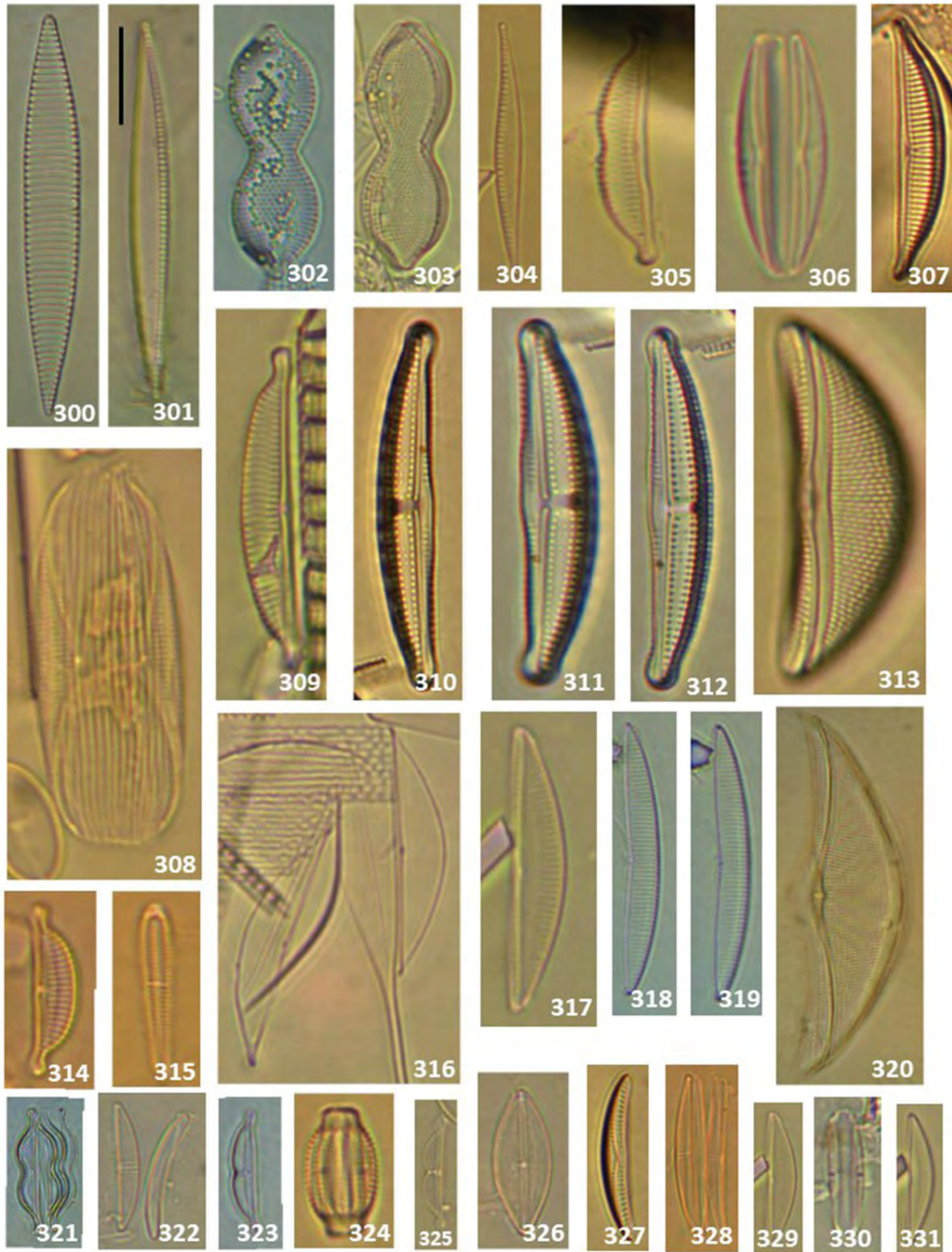
Figures 220-248. 220. *Diploneis crabro* f. *dirhombus*; 221. *Diploneis chersonensis*; 222, 223. *Diploneis crabro*; 224, 225. *Caloneis maxima* var. *excentrica*; 226. *Caloneis elongata*; 227, 241, 244. *Caloneis linearis*; 228. *Diploneis smithii*; 229, 230. *Caloneis* sp. 4; 231, 232. *Caloneis* sp. 3; 233, 239. *Diploneis vacillans* var. *renitens*; 234, 242. *Caloneis* sp. 5; 235, 243. *Caloneis* sp. 1; 236. *Caloneis latiuscula* f. *minor*; 237, 238. *Diploneis vacillans* var. *vacillans*; 240. *Mastogloia crucicula*; 245. *Diploneis litoralis* var. *clathrata*; 246. *Diploneis litoralis*; 247. *Caloneis* sp. 2; 248. *Diploneis parca*.



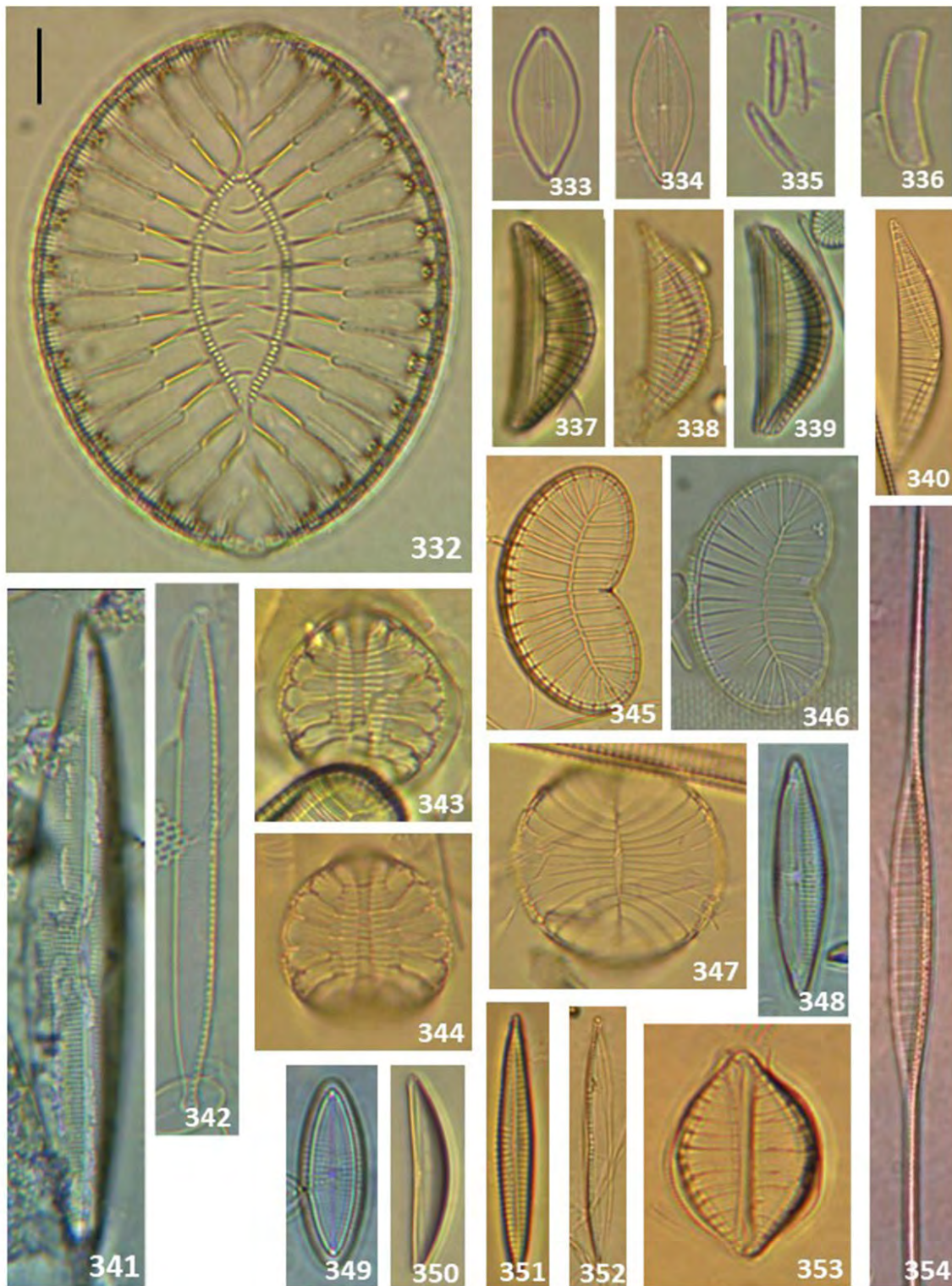
Figures 249-277. 249, 269. *Navicula longa*; 250, 251. *Navicula johanrossii*; 252. *Navicula leptoloba*; 253, 254, 274, 275. *Navicula zosteretii*; 255, 256. *Gyrosigma tenuissimum*; 257. *Haslea nautica*; 258, 259. *Tryblionella marginulata* var. *didyma*; 260, 261. *Psammodictyon constrictum*; 262, 265, 266, 268. *Cymbellonitzschia* sp. 2; 263, 267. *Cymbellonitzschia banzuensis*; 264. *Nitzschia persuadens*; 270, 271. *Hantzschia amphioxys*; 272. *Navicula halophila*; 273. *Navicula cincta*; 276. *Parlibellus* cf. *phoebeae*; 277. *Haslea* cf. *howeana*.



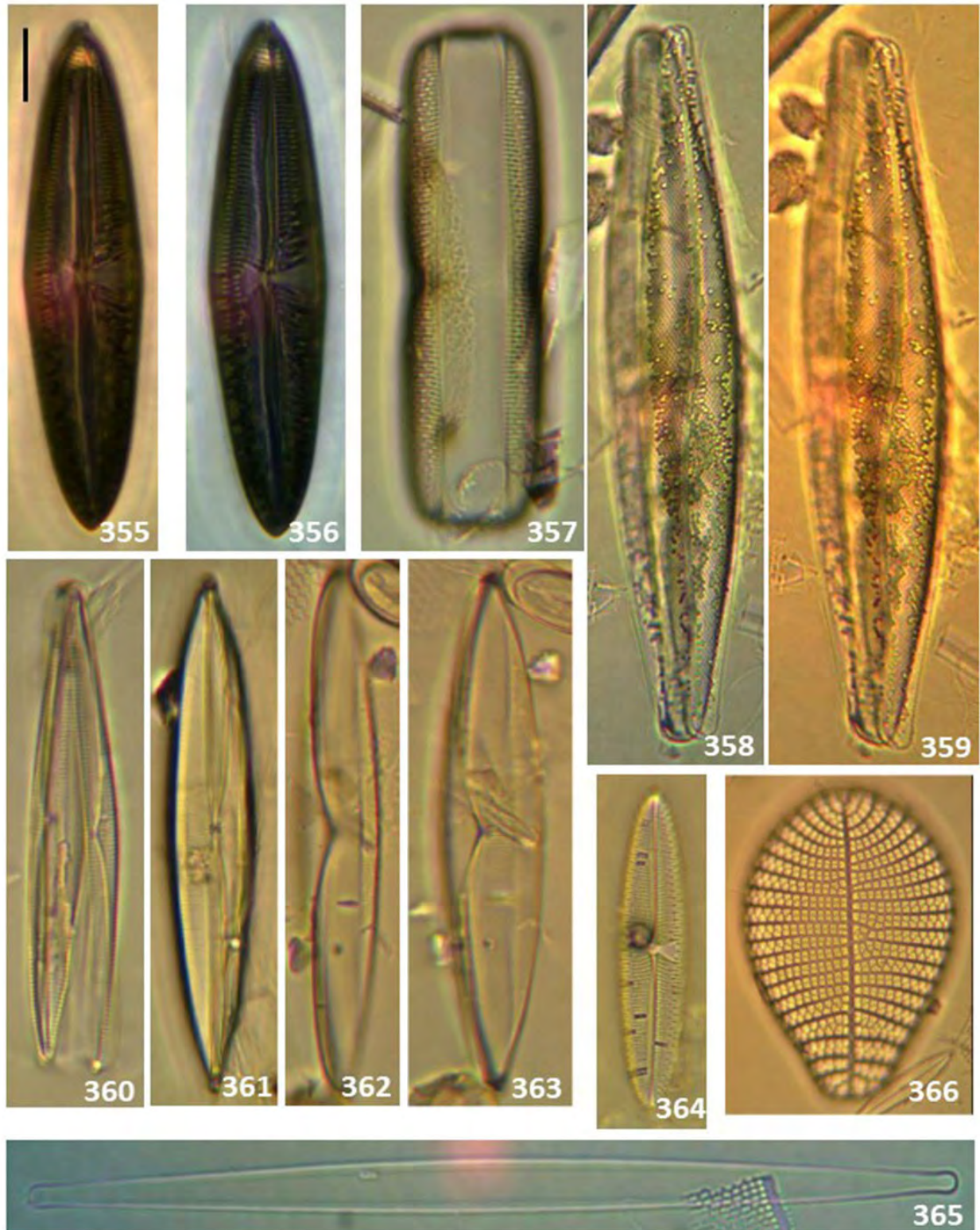
Figures 278-299. 278, 280. *Tryblionella coarctata*; 279. *Psammodictyon constrictum* var.; 281, 287. *Psammodictyon constrictum*; 282, 296. *Nitzschia subacuta*; 283. *Nitzschia sicula*; 284. *Nitzschia lanceolata*; 285, 286. *Tryblionella marginulata* var. *didyma*; 288, 294. *Nitzschia lanceolata* var. *minor*; 289. *Hyalosynedra laevigata*; 290. *Synedra commutata*; 291-293. *Nitzschia bicapitata*; 291a. *Nitzschia* cf. *panduriformis* var. *continua*; 292a. *Nitzschia persuadens*; 293a, 294a. *Nitzschia dissipata*; 295. *Diploneis vacillans* var. *renitens*; 297. *Nitzschia confinis*; 298, 299. *Cyclophora tenuis*.



Figures 300-331. 300. *Nitzschia sicula*; 301. *Nitzschia sigma*; 302, 303. *Nitzschia bombiformis*; 304. *Nitzschia sigma* var.; 305. *Halamphora angularis*; 306. *Amphora exilitata*; 307. *Halamphora coffeaeformis*; 308. *Amphora lineolata*; 309. *Halamphora dusenii*; 310-312. *Amphora maletracta* var. *constricta*; 313. *Amphora proteus*; 314, 324. *Halamphora turgida*; 315. *Gomphonemopsis pseudexigua*; 316. *Amphora pseudohyalina*; 317, 329, 331. *Seminavis delicatula*; 318, 319. *Amphora* sp.1; 320. *Amphora rhombica*; 321, 323, 325. *Amphora bigibba*; 322, 328. *Halamphora* cf. *staurophora*; 326. *Mastogloia borneensis*; 327. *Amphora proteus* var. *contigua*; 330. *Halamphora staurophora*.



Figures 332-354. 332. *Campylodiscus neofastuosus*; 333, 334. *Mastogloia borneensis*; 335. *Catenula* sp.; 336. *Rhoicosphenia* cf. *genuflexa*; 337, 339. *Epithemia pacifica*; 338. *Rhopalodia gibberula*; 340. *Rhopalodia* sp.; 341. *Navicula libellus*; 342. *Nitzschia tubicola*; 343, 344. *Campylodiscus thuretii*; 345, 346. *Plagiodiscus nervatus*; 347. *Campylodiscus decorus*; 348. *Parlibellus cruciculoides*; 349. *Navicula comoides*; 350. *Seminavis* sp.; 351. *Navicula leptoloba*; 352. *Nitzschia pellucida*; 353. *Rhopalodia gibberula* var. *producta*; 354. *Nitzschia longissima* var. *costata*.



Figures 355-366. 355, 356. *Trachyneis aspera* var. *oblonga*; 357, 364. *Trachyneis aspera*; 358, 359. *Pleurosigma distinguendum*; 360. *Tropidoneis* sp.; 361. *Tropidoneis pusilla*; 362, 363. *Tropidoneis vitrea*; 365. *Ardissonaea* cf. *crystallina*; 366. *Podocystis americana*.

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